Short Communication

Efficacy and safety of plerixafor in pediatric cancer patients undergoing peripheral blood stem cell harvest for autologous hematopoietic stem cell transplant

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Abstract

Plerixafor for peripheral blood hematopoietic stem cell (PB HSC) mobilization in children undergoing autologous hematopoietic stem cell transplantation is primarily used following failure of the initial mobilization attempt. Data on plerixafor use in pediatric patients are limited.

This retrospective study conducted at a single tertiary care center in India, details the efficacy and safety of plerixafor for 10 children with relapsed/refractory solid tumors or lymphomas. High risk neuroblastomas (HR NB) underwent autologous HSCT as part of consolidation. Plerixafor was administered at a dose of 240 μg/kg body weight of the recipient, subcutaneously, approximately 11-12 h prior to harvest. Ten patients (eight males, two females), with a median age of 8 years (range 2-18 years), received plerixafor prior to PB HSC harvest. All patients were administered granulocyte colony stimulating factor (GCSF) before the administration of plerixafor. The median CD 34 count for all patients pre-plerixafor was 29/μL, nine patients exhibited higher CD 34 post plerixafor (median of 148/μL). In nine patients, the values of the CD 34 count and total leukocyte count (TLC) of the harvested product were available, and in all cases, we achieved a good yield. All patients in this study were heavily chemotherapy pre-treated, and the use of plerixafor resulted in a satisfactory yield of peripheral blood stem cells. No side effects were observed.

Key words plerixafor, pediatrics, autologous transplant

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Introduction

Autologous peripheral blood stem cell transplantation (A-PBSCT) has become the standard of care for many malignancies and its use is increasing in non-malignant conditions, such as autoimmune diseases 1. For optimal outcomes post-A-PBSCT, it is imperative that an adequate number of hematopoietic stem cells (HSC) are infused. Optimal outcomes are achieved if more than 2.5 million HSC per kg body weight is administered post high-dose conditioning chemotherapy 2.

Few strategies can be used for the mobilization of HSC into the peripheral blood to allow for collection. The most common method involves the use of granulocyte colony-stimulating factor (GCSF) alone or following the administration of a high-dose of cyclophosphamide or chemotherapy. However, in patients that have undergone significant chemotherapy, these methods may result in mobilization failure. Furthermore, in such patients, repeat apheresis may be required, causing delays in treatment and additional burdens on healthcare resources. It may also rarely necessitate bone marrow harvest, which is a more invasive procedure requiring the administration of general anesthesia 3.

Plerixafor is an antagonist of the CXCR4 chemokine receptor, and its use in combination with GCSF to aid HSC mobilization has been approved in adults in the United States of America and Europe. Plerixafor reversibly inhibits the interaction of the CXCR4 receptor, present on the surface of stem cells, with the stromal cells in the bone marrow niches that carry the CXCL2 ligand. This prevents the attachment of HSC to stromal
cells, allowing them to enter peripheral blood circulation\(^1\). The MOZAIC study was a phase I/II study conducted in children, which established the dose of plerixafor for use in phase II of the trial (n=45; plerixafor arm=30, standard arm=15). It was shown to be safe and effective for HSC mobilization in children\(^2\). The use of plerixafor in children, remains understudied and further evidence is required.

We performed this retrospective analysis of 10 pediatric patients diagnosed with lymphomas and solid tumors, in whom plerixafor was administered for the mobilization of HSC to enable harvesting. Here, we investigated the efficacy and safety of plerixafor administration in these children.

**Methods**

The records of patients who underwent A-PBSCT as part of their treatment were analyzed. The details of the patients who received plerixafor for the mobilization of HSC were collected. The demographic details, diagnosis, previous chemotherapy administered, doses of GCSF administered, CD 34 and total leukocyte count (TLC) pre- and post-harvest along with CD 34 and TLC of the harvested product were recorded. GCSF was administered subcutaneously at a dose of 5 µg/kg/dose twice a day or 10 µg/kg/dose once a day. Plain GCSF was used in all cases. The number of days GCSF was administered varied between patients, as admission for the apheresis procedure depended on the bed availability. Plerixafor was administered to the patient 11-12 hours prior to harvest at a dose of 240 µg/kg. Plerixafor was administered if the CD 34 count was less than 50/µL. We chose a CD 34 count of 50/µL, based on previous experience when apheresis was performed on patients with lower CD 34 counts as it yielded suboptimal doses of stem cells in the final product. In instances when the CD 34 count was not available on the day prior to the harvest, plerixafor was administered irrespective of the count. Enumeration of CD34 percentage was performed using flow cytometry. The absolute CD 34 count was obtained using TLC and CD 34 percentage.

Adverse effects after the administration of plerixafor were recorded, including allergic and hypersensitivity reactions, thrombocytopenia, respiratory difficulties, bone pain, and splenomegaly. Vital signs and laboratory parameters were recorded for all patients at regular intervals after the administration of plerixafor.

The data were analyzed using descriptive statistics. Continuous variables were analyzed using the mean, standard deviation (SD), median (range), and categorical variables were summarized using numbers and proportions.

**Results**

Ten patients received plerixafor before the peripheral blood hematopoietic stem cell (PB HSC) harvest was performed. There were 8 male and 2 female patients, with a median age of 8 y (2-18 y).

Four patients had relapsed/refractory Hodgkin lymphoma (2 relapsed/2 refractory), one patient had refractory diffuse large B-cell lymphoma (DLBCL), three patients had high-risk neuroblastoma, and two patients had relapsed retinoblastoma. The mean weight of the patients was 30.9 ± 17.6 kg. Patients were administered either 5 µg/kg twice a day or 10 µg/kg once a day of GCSF. The patients received variable cumulative doses of GSCF before receiving plerixafor prior to admission for apheresis, and the mean dose given was 51.0 ± 13.3 µg/kg.

The peripheral blood CD 34 count on the day prior to the harvest was less than 50/µL in nine cases, and in one child (patient no. 6), the CD 34 count was not available.

The median CD 34 count for all patients on the day prior to harvest was 29/µL (3-95/µL) and the corresponding TLC was 40,991/µL (17,400-65,830/µL). Following plerixafor administration, there was a significant increase in the CD 34 count and TLC in all patients, except one (patient no. 5). In the nine patients in whom there was a rise in counts after plerixafor, the CD 34 count post plerixafor increased to a median of 148/µL (26-458/µL) and the TLC rose to a median of 53,195/µL (31,390-95,690/µL). In nine cases, we were able to obtain the CD 34 count of the harvested product, the median CD 34 count was 2,606/µL (850-6,047/µL). The stem cell doses collected for all patients are shown in Table 1.

None of the patients experienced any side effects related to plerixafor.

**Discussion**

In our retrospective study, we found that the use of plerixafor post-GCSF led to an increase in the CD 34 counts in the peripheral blood, allowing for an improved yield of CD 34 positive HSC. The use of plerixafor was not associated with any adverse effects.

In pediatric autologous HSCT, a good stem cell dose is important. A stem cell count of at least 2-5 million cells per kg is recommended. As higher stem cell doses are associated with faster engraftment and better overall outcomes\(^3\). In cases of tandem transplants, larger numbers of stem cells need to be collected for use in more than one rescue after high-dose chemotherapy.

Mobilization using GCSF has been predominantly used for stem cell harvest, but it carries a 5-30% risk of
failure, and this could be more in patients treated with multiagent chemotherapy. Poor mobilization may lead to the abandonment of A-PBSCT as a treatment option. Remobilization carries the risk of morbidity, inconveniences for the patient, and increased utilization of resources. Plerixafor alone or in combination with GCSF has been reported to significantly increase the number of PB HSC cells and is being increasingly used as a modality to avoid mobilization failure. However, evidence in the pediatric population is still emerging. Teusink et al (2016) have reported the use of plerixafor in 16 children undergoing A-PBSCT, following failed initial mobilization attempts, and demonstrated that 14 patients were able to achieve optimal CD 34 counts for autologous HSCT. The phase I/II MOZAIC trial in children between 1–18 years with solid tumors, lymphomas, and brain tumors showed that with plerixafor and standard mobilization, the proportion of patients with doubled PB CD 34 HSC counts significantly increased when compared to standard mobilization alone. In 18 patients with solid tumors, plerixafor was found to be effective in increasing PB HSC counts, permitting a successful harvest. In this study, the use of plerixafor led to an increase in PB HSC counts in nine out of ten children. In eight children who received plerixafor, one dose was sufficient to achieve an acceptable HSC yield for A-PBSCT, and a second apheresis procedure was avoided.

Maschan et al. reported mild side effects in approximately 25% of children aged 1-18 years receiving plerixafor. Sevilla et al. also found mild adverse effects in two of eight patients. Serious side effects are rare, but some, such as spontaneous pneumomediastinum after the use of plerixafor, have been described in the literature. None of the patients in this study experienced any side effects and plerixafor was well tolerated. A low rate of adverse effects has been reported in 18 Japanese children who received plerixafor. The MOZAIC study also demonstrated that plerixafor was safe for use in children, and all the adverse effects were attributable to mobilizing chemotherapy.

The findings of recent pediatric studies on the use of plerixafor is presented in Table 2.

### Table 1. Clinical details for all patients

<table>
<thead>
<tr>
<th>S I</th>
<th>A g e (years)</th>
<th>Sex</th>
<th>Weight (kg)</th>
<th>Diagnosis</th>
<th>Cumulative GCSF dose (in μg/kg) before plerixafor</th>
<th>CD 34/TLC before plerixafor (per μL of patients blood volume)</th>
<th>No of doses of plerixafor</th>
<th>CD 34/TLC post plerixafor (per μL of patients blood volume)</th>
<th>CD34/TLC harvested of product (per μL of harvested product)</th>
<th>Stem cell dose/kg of recipient weight collected (× 10^5/kg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>7</td>
<td>Male</td>
<td>17.3</td>
<td>Refractory HL</td>
<td>60</td>
<td>23/24,710</td>
<td>2 (2 harvests)</td>
<td>26/31,390</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>2</td>
<td>4</td>
<td>Male</td>
<td>15</td>
<td>HR NB</td>
<td>50</td>
<td>20/50,410</td>
<td>2 (2 harvests)</td>
<td>65/53,990</td>
<td>1,062/212,390</td>
<td>9.6</td>
</tr>
<tr>
<td>3</td>
<td>9</td>
<td>Male</td>
<td>34</td>
<td>Relapsed HL</td>
<td>40</td>
<td>19/65,830</td>
<td>1</td>
<td>34/72,410</td>
<td>2,362/218,700</td>
<td>14.6</td>
</tr>
<tr>
<td>4</td>
<td>2</td>
<td>Male</td>
<td>13.4</td>
<td>HR NB</td>
<td>70</td>
<td>30/17,400</td>
<td>1</td>
<td>458/53,890</td>
<td>3,162/263,500</td>
<td>15.2</td>
</tr>
<tr>
<td>5</td>
<td>17</td>
<td>Male</td>
<td>70</td>
<td>Refractory HL</td>
<td>30</td>
<td>36/56,030</td>
<td>1</td>
<td>33/32,700</td>
<td>1,257/262,000</td>
<td>6.0</td>
</tr>
<tr>
<td>6</td>
<td>6</td>
<td>Male</td>
<td>20</td>
<td>Relapsed metastatic RB</td>
<td>55</td>
<td>95/51,420</td>
<td>1</td>
<td>295/68,900</td>
<td>5,279/707,680</td>
<td>31.1</td>
</tr>
<tr>
<td>7</td>
<td>18</td>
<td>Male</td>
<td>51</td>
<td>Relapsed HL</td>
<td>75</td>
<td>50/22,820</td>
<td>1</td>
<td>151/42,000</td>
<td>3,418/280,200</td>
<td>16.7</td>
</tr>
<tr>
<td>8</td>
<td>14</td>
<td>Female</td>
<td>38</td>
<td>Refractory DBLCL</td>
<td>45</td>
<td>42/33,730</td>
<td>1</td>
<td>215/52,500</td>
<td>850/250,000</td>
<td>4.7</td>
</tr>
<tr>
<td>9</td>
<td>11</td>
<td>Female</td>
<td>35</td>
<td>Relapsed metastatic RB</td>
<td>40</td>
<td>3/59,220</td>
<td>1</td>
<td>287/95,690</td>
<td>6,047/295,000</td>
<td>31.9</td>
</tr>
<tr>
<td>10</td>
<td>3</td>
<td>Male</td>
<td>16</td>
<td>HR NB</td>
<td>45</td>
<td>28/28,340</td>
<td>1</td>
<td>145/42,700</td>
<td>2,606/357,000</td>
<td>24.4</td>
</tr>
</tbody>
</table>

GCSF, granulocyte colony stimulating factor; TLC, total leukocyte count; HL, Hodgkin lymphoma; HR NB, high risk neuroblastoma; RB, retinoblastoma; DBLCL, diffuse large B cell lymphoma
Table 2. Recent pediatric studies documenting the use of plerixafor

<table>
<thead>
<tr>
<th>Author/year/reference</th>
<th>Number of patients (n)</th>
<th>Age (in years)</th>
<th>Mobilization strategy</th>
<th>Dose of plerixafor</th>
<th>Conclusion</th>
</tr>
</thead>
<tbody>
<tr>
<td>Goto et al 2022</td>
<td>18</td>
<td>1-13</td>
<td>Chemotherapy followed by GCSF and Plerixafor</td>
<td>0.24 mg/kg daily</td>
<td>66.7% achieved target CD34 cell count and pyrexia was the commonest adverse effect</td>
</tr>
<tr>
<td>Morland et al 2020</td>
<td>45 (30 received plerixafor and 15 received standard treatment) &lt;18 y</td>
<td>GCSF and plerixafor</td>
<td>0.24 mg/kg 8-12 h prior to apheresis</td>
<td>80% patients in plerixafor arm achieved doubling of CD34 compared to 28.6% in standard arm</td>
<td></td>
</tr>
<tr>
<td>Teusnik et al 2016</td>
<td>16</td>
<td>0.7-15.4</td>
<td>GCSF and plerixafor</td>
<td>0.24 mg/kg</td>
<td>An adequate number of CD34+ cells were obtained in 14 of 16 patients (87.5%).</td>
</tr>
<tr>
<td>Maschan et al 2015</td>
<td>33</td>
<td>1-18 y</td>
<td>GCSF and plerixafor</td>
<td>0.24 mg/kg in 30 patients and 0.3 µg/kg in 3 patients</td>
<td>31 of 33 patients mobilized successfully</td>
</tr>
<tr>
<td>Sevilla et al 2012</td>
<td>8</td>
<td>6-18 y</td>
<td>GCSF and plerixafor in 5 GCSF and plerixafor with chemotherapy in 3</td>
<td>0.24 mg/kg</td>
<td>Peripheral blood progenitor cell priming was improved with plerixafor in 6 of 8 patients.</td>
</tr>
</tbody>
</table>

GCSF, granulocyte colony stimulating factor

Author Contributions

AKG drafted the original manuscript. AKG, JPM, HCP, PC and RS contributed to patient care. All authors revised the manuscript. All authors approved the final manuscript.

Consent

Informed consent was obtained by all participants in this study.

Conflicts of Interest

The authors declare no conflict of interest. Disclosure forms provided by the authors are available on the website.

References


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