Original Article



A Novel Hematopoietic Progenitor Cell Mobilization Regimen Comprising Bortezomib, G-CSF, and Preemptive Plerixafor for Multiple Myeloma

Kazuki Tanimoto, Keiji Sakamoto, Ichiro Kawano, Ikumi Yamanaka, Hideho Henzan

Department of Hematology and Oncology, Japanese Red Cross Fukuoka Hospital, Fukuoka, Japan

Abstract

Adequate hematopoietic progenitor cell collection is critical for autologous peripheral blood stem cell transplantation. Conventionally, patients with multiple myeloma are treated with high-dose cyclophosphamide and granulocyte colony-stimulating factor (G-CSF) or G-CSF alone to mobilize their peripheral blood stem cells. However, some patients exhibit insufficient stem cell recruitment in response to these regimens. Recently, plerixafor has been approved for coverage by insurance in Japan. Combination treatment with plerixafor and G-CSF is now a standard procedure. In addition, treatment with bortezomib and G-CSF results in efficient stem cell recruitment. On the basis of the results from mouse studies, we hypothesized that combination treatment with bortezomib ensures efficient mobilization and mediates in vivo purging of malignant cells. Therefore, we administered a regimen of bortezomib, G-CSF, and preemptive plerixafor to 10 patients with multiple myeloma, and analyzed its efficacy and safety. The median patient age was 68 years. We collected CD34-positive cells (median: 4.9×10^6 /kg) in a single session of apheresis from all patients. We observed no obvious myeloma cell contamination in the collected product or serious toxicity during treatment and collection. After collection, we performed autologous peripheral blood stem cell transplantation and confirmed engraftment in all patients (median: day 10). We found that the regimen is safe and reliably facilitated the collection of sufficient autologous peripheral blood stem cells by apheresis from all patients in a single day. Despite the small patient group size, we conclude that the regimen is promising for safe and efficient collection of peripheral blood stem cells for autologous transplantation in patients with multiple myeloma.

Key words: multiple myeloma, peripheral blood mobilization, bortezomib, plerixafor, granulocyte colony-stimulating factor

Submitted May 25, 2020; Accepted July 29, 2020; Published online October 9, 2020; Issued online November 25, 2020 Correspondence: Kazuki Tanimoto, Department of Hematology and Oncology, Japanese Red Cross Fukuoka Hospital, 3-1-1 Okusu, Minami-ku, Fukuoka-city, Fukuoka, 810-8555, Japan, E-mail: kaztanimo@fukuoka-med.jrc.or.jp

Introduction

For patients with transplant-eligible multiple myeloma (MM), high-dose chemotherapy with autologous hematopoietic stem cell transplantation results in a better progression-free survival than conventional chemotherapy, as described in clinical trials¹. Conventionally, a regimen consisting of high-dose cyclophosphamide followed by granulocyte colony-stimulating factor (G-CSF) or G-CSF alone has been used to recruit peripheral blood stem cells (PBSCs) in patients with MM. However, some patients do not mobilize sufficient hematopoietic progenitor cells (HPCs) for transplantation. Musto *et al.* reported predictors for unsuccessful or sub-optimal HPC mobilization, which include advanced age, lenalidomide treatment, and severe toxicity in response to high-dose cyclophosphamide². The use of plerixafor in combination with G-CSF improves mobilization compared with G-CSF alone or in combination with cyclophosphamide, and the plerixafor combination is recommended for a standard collection method³⁻⁵. However, even after using plerixafor, some patients still show poor mobilization. Furthermore, the possibility of tumor cell contamination of plerixafor-mobilized HPCs cannot be completely ruled out^{6,7}. In fact, Morris *et al.* reported that the cumulative incidence of relapse after transplantation was higher in the plerixa-for cohort than in the other regimen cohorts⁸.

A previous study reported that stem cell recruitment

with bortezomib and G-CSF is efficient⁹. In this study, bortezomib was administered on days -11 and -8, followed by G-CSF on days -4 to -1, before HPC collection. The median circulating CD34 + cell count on mobilization day before plerixafor administration was 25.4 cells/ μ L, and two patients (11%) in the study received plerixafor⁹. The safety and feasibility of bortezomib in combination with G-CSF for PBSC collection have been demonstrated in a phase I study¹⁰. In this study, G-CSF was administered for 5 days, and on the evening of day 4, a single dose of bortezomib was administered. The median circulating CD34+ cell count on mobilization day was 32 cells/ μ L without plerixafor administration¹⁰. Although there is a difference in the administration schedule of bortezomib, these two studies reveal that bortezomib in combination with G-CSF effectively mobilizes PBSCs. Combination therapy with bortezomib is expected to ensure therapeutic effects against MM and mediate in vivo purging due to the duration of proteasome inhibition in the peripheral blood^{11,12}. Moreover, in a mouse model, plerixafor administration after bortezomib further enhanced HPC mobilization compared with the administration of either agent alone¹¹.

We hypothesized that the addition of bortezomib to the standard plerixafor and G-CSF regimen would result in higher mobilization than the standard regimen alone, with the additional benefits of anti-myeloma and *in vivo* tumor purging effects. The enhanced mobilizing activity and tumor-purging effects of bortezomib are highly desirable for patients with MM. Based on these observations, the aim of our pilot study was to determine the safety and efficacy of a unique autologous PBSC collection regimen, comprising plerixafor administration, following treatment with bortezomib and G-CSF for blood stem cell mobilization in patients with MM.

Patients and Methods

We obtained Institutional Review Board approval from the Japanese Red Cross Fukuoka Hospital. All patients provided written informed consent in accordance with the Declaration of Helsinki. We analyzed the clinical outcomes in adult patients requiring their first PBSC transplant for newly diagnosed MM between April 2017 and March 2019 in our hospital.

The PBSC collection regimen is shown in **Figure 1**. The patients were then subcutaneously administered bortezomib (1.3 mg/m^2) on days 1 and 4, G-CSF $(10 \mu \text{g/kg})$ for 5 consecutive days, and preemptive plerixafor (0.24 mg/kg) on the evening of day 4. We determined circulating peripheral blood CD34-positive cell counts on the morning of days 4 and 5. If the count was >20 cells/ μ L on the morning of day 4, we suspended the preemptive plerixafor (0.24 mg/kg) treatment. The collection

| | | Day | 1 | 2 | 3 | 4 | 5 |
|---|-----------------------|-----|----|----|----|----|---|
| Bortezomib | 1.3 mg/m ² | SC | ₽ | | | - | |
| G-CSF | 10 µg/kg | SC | 11 | 11 | 11 | 11 | Ţ |
| Plerixafor | 0.24 mg/kg | SC | | | | (| |
| Apheresis | | | | | | | Ŧ |
| Peripheral blood CD34-positive cell count | | | | | | 1 | t |

Figure 1. Regimen scheme

 (\downarrow) If the peripheral blood CD34-positive cell count was>20 cells/ μ L on the morning of day 4, preemptive plerixafor (0.24 mg/kg) treatment was suspended.

sc, subculaneous.

goal was 2.0×10^6 CD34-positive cells/kg, based on the recommended adequate dose for one autologous hematopoietic stem cell transplantation^{13.} If the goal was not met after 1 day of apheresis, we planned repeat G-CSF administration and apheresis.

We performed autologous PBSC collection on day 5 using a standard procedure. After adequate cell collection, we performed autologous PBSC transplantation after conditioning with melphalan.

Table 1 shows the patient characteristics. Six patients were men (60%), with the median age of 68 (range: 45-74) years. Six patients had Bence Jones type M proteins. Nine patients received an induction therapy of bortezomib-lenalidomide-dexamethasone. Nine patients received four or more prior cycles of induction treatment that included lenalidomide. Four patients achieved a stringent complete response (sCR), one achieved a complete response (CR), four patients showed a very good partial response (VGPR), and one patient showed a partial response (PR).

Results

The results of the cell collection procedures are shown in **Table 1**. The median CD34-positive cell yield was 4.9×10^6 cells/kg (range: $2.3-9.8 \times 10^6$ cells/kg). We acquired an adequate amount of cell from all patients (100%) with a single session of apheresis. We collected a sufficient amount of cells without plerixafor from 2 of the 10 patients (2.93 and 3.24×10^6 /kg) because their CD34-positive peripheral blood cell count on the morning of day 4 had reached adequate levels before plerixafor administration.

Figure 2 shows the circulating peripheral blood CD34positive cell count in patients on the morning of days 4 and 5. The median count was 13.9 (range: 3.0-39.9) cells/ μ L on the morning of day 4. The median count was 48.9 (range: 24.1-110.5) cells/ μ L on the morning of day 5, measured 15 to 18 h post bortezomib treatment on day 4, 8 h post plerixafor treatment on day 4, and before

| Age | Sex | Induction | Response at collection | Day 4 CD34+ ^a | Day 5 CD34+ ^a | Day 5 CD34+ yield ^b | Days of apheresis | Plerixafor |
|-----|-----|--------------|------------------------|-----------------------------|-----------------------------|--------------------------------------|-------------------|------------|
| 66 | М | VRD 4 course | sCR | 21.99 | 100.61 | 9.79 | 1 | + |
| 58 | F | VCD 4 course | VGPR | 18.97 | 48.06 | 3.48 | 1 | + |
| 74 | Μ | VRD 5 course | VGPR | 3.76 | 27.7 | 4.09 | 1 | + |
| 73 | Μ | RD 4 course | VGPR | 16.95 | 110.54 | 5.68 | 1 | + |
| 68 | F | VRD 4 course | sCR | 18.89 | 24.14 | 2.93 | 1 | — |
| 45 | Μ | VRD 4 course | sCR | 39.92 | 50.01 | 3.24 | 1 | — |
| 73 | Μ | VRD 4 course | PR | 4.69 | 26.8 | 2.27 | 1 | + |
| 65 | F | VRD 4 course | VGPR | 3.93 | 45.24 | 6.20 | 1 | + |
| 70 | F | VRD 4 course | sCR | 3.05 | 49.67 | 5.70 | 1 | + |
| 58 | М | VRD 4 course | CR | 10.91 | 68.43 | 7.22 | 1 | + |

Table 1. Patient characteristics

F, female; M, male; VRD, bortezomib, lenalidomide, and dexamethasone; VCD, bortezomib, cyclophosphamide, and dexamethasone; RD, lenalidomide and dexamethasone; sCR, stringent complete response; VGPR, very good partial response; CR, complete response; PR, partial response; CD34+, CD34-positive cell.

^a cells/ μ L ^b 10⁶ colls/k

^o 10⁶ cells/kg



Figure 2. Circulating peripheral blood CD34-positive cell counts on the morning of days 4 and 5 (A) Peripheral blood CD34-positive cell count on days 4 and 5. The data are presented as mean±standard error.(B) Peripheral blood CD34-positive cell counts in each patient.

G-CSF administration on day 5. The median CD34-positive cell counts on days 4 and 5 were significantly different as assessed using the *t*-test (P = 0.0008; **Figure 2A**). The cell counts on the morning of days 4 and 5 revealed that the circulating peripheral blood CD34-positive cells were efficiently mobilized on the last day of the regimen. The median increase in the circulating CD34-positive peripheral blood cell count from the morning of day 4 to the morning of day 5 was 35.2 (range: 5.25-93.59) cells/ μ L, showing almost a 6-fold increase (range: 1.28- to 7.37-fold). This increase was higher in the patients who

received plerixafor than in those who did not (**Figure 2B**).

Table 2 shows the adverse events in all patients. We evaluated the grade of the adverse events using Common Terminology Criteria for Adverse Events version 4 (https://ctep.cancer.gov/protocolDevelopment/electr onic_applications/ctc.htm). All patients had mild toxicities, no serious toxicities were reported during treatment or collection. The most common adverse event related to mobilization was back or musculoskeletal pain (50%). There were no grade 3 or higher toxicities, death, or

Table 2. Adverse events

| | Number of events (%) | Number of grade 3 and 4 events |
|--|----------------------|--------------------------------|
| Any class | 10 (100%) | 0 |
| Infections and infestations | 0 | 0 |
| Nervous system disorders | 1 (10%) | 0 |
| Neuropathy, peripheral disorders | 1 (10%) | 0 |
| Cardiac disorders | 0 | 0 |
| Respiratory disorders | 0 | 0 |
| Gastrointestinal disorders | 5 (50%) | 0 |
| Diarrhea | 3 (30%) | 0 |
| Abdominal discomfort | 1 (10%) | 0 |
| Constipation | 4 (40%) | 0 |
| Vomiting | 3 (30%) | 0 |
| Hepatobiliary disorders | 0 | 0 |
| Skin and subcutaneous tissue disorders | 0 | 0 |
| Musculoskeletal and connective tissue disorders | 5 (50%) | 0 |
| Back pain | 5 (50%) | 0 |
| Musculoskeletal pain | 2 (20%) | 0 |
| General disorders and administration site conditions | 2 (20%) | 0 |
| Fatigue | 1 (10%) | 0 |
| Fever | 2 (20%) | 0 |
| Investigations | 1 | 0 (100%) |
| Elevation in LDH (>1.5N) | 1 | 0 (100%) |
| Elevation in CRP (>1.5N) | | 4 (40%) |

LDH, lactate dehydrogenase; CRP, C-reactive protein; >1.5N, 1.5 times more than the normal range.

treatment discontinuation due to adverse events.

After conditioning with melphalan, all patients underwent autologous stem cell transplantation with their total cell products. After autologous stem cell transplantation, we confirmed rapid engraftment in all patients. The median days post-transplant to reach a neutrophil count of $>500/\mu$ L and platelet count of $>20,000/\mu$ L were 10 (range: day 9-11) and 12 (range: day 10-21), respectively.

Discussion

The consensus of the American Society for Blood and Marrow Transplantation is upfront plerixafor is a suitable treatment option for all patients¹³. We demonstrated the efficacy of PBSC collection with preemptive use of plerixafor following bortezomib and G-CSF administration. The rate of successful HPC collection using our regimen was superior to that using G-CSF and plerixafor in patients with MM14. With strategies employed in previous studies, such as chemotherapy and G-CSF and G-CSF alone, a significant number of patients (15%-30%) failed to mobilize an adequate number of stem cells^{15,16}. With G-CSF + plerixafor therapy, the proportion of patients from whom $> 2 \times 10^6$ CD34-positive cells/kg were collected in 4 or fewer apheresis sessions was 88.3%¹⁴. Similarly, two other studies reported that for adequate cell collection, the median number of aphereses

was 2 (range: 1-4)^{3,4}. Moreover, in a comparative cohort of Japanese patients, Yogo et al. reported that 17% of 63.8% patients who received lenalidomide did not yield more than 2.0×10^6 CD34 + cells/kg with a single apheresis session¹⁷. These revealed that the frequency of insufficient cell collection using G-CSF + plerixafor was 10%-20%. However, using our regimen, we collected sufficient amount of cells from all 10 (100%) patients with a single session of apheresis. The superior mobilization achieved with our regimen can be attributed to bortezomib administration. Bortezomib has been shown to function in the bone marrow microenvironment to inhibit the adhesion of HPC to bone marrow stromal cells^{18,19}. Furthermore, HPCs were more mobilized in mice co-administered bortezomib and plerixafor¹¹. Therefore, we expected that the administration of plerixafor and G-CSF simultaneously could mobilize more CD34-positive cells. To the best of our knowledge, only a few studies have shown that the simultaneous administration of these two drugs enhanced mobilization effect. Therefore, we added bortezomib on days 1 and 4 to a regimen comprising plerixafor and G-CSF, according to the convenience based on the logistics of the clinic.

A previous study reported that age, baseline low peripheral blood cell count, lenalidomide use, and hematological toxicity during induction were potential factors associated with poor mobilization under cyclophosphamide treatment². In this study, the amount of CD34 + PBSC cells (<CD34 + 2.0 × 10⁶ cells/kg) from patients collected was not sufficient, 15.54% of whom were over 60 years old and 15.17% had a history of lenalidomide use. Considering that our patients were elderly individuals and had a history of treatment with four or more courses of lenalidomide, it is surprising that we were able to obtain enough CD34-positive cells in a single session of apheresis. The ability to collect a sufficient amount of cells in a single apheresis session may provide patients a sense of security and reduce the number of collections. We speculate that even if this regimen is used in a higher number of patients, it is likely that a single harvest will be sufficient, because our patients in this trial had a relatively higher risk of collection failure.

We suggest that the combination therapy of bortezomib and plerixafor might have enhanced therapeutic effects against MM. A recent phase I/II trial revealed that plerixafor in combination with bortezomib was safe, and yielded a high objective response rate even in patients with relapsed/refractory MM²⁰. Moreover, we expected to observe a reduction in the number of circulating myeloma cells with bortezomib combination regimen based on long-lasting *in vivo* purging observed in mice¹². We chose to add bortezomib on both days 1 and 4 due to its direct anti-myeloma effects.

The consensus of the American Society for Blood and Marrow Transplantation is the preemptive use of plerixafor based on peripheral blood CD34-positive cell measurements is reasonable¹³. We examined circulating peripheral blood CD34-positive cells on the morning of day 4. Yogo et al. reported PBSC mobilization in 67 patients who received G-CSF alone and G-CSF + plerixafor. The median CD34-positive cell count on days 4 and 5 in the G-CSF + plerixafor group was 6 and 38 cells/ μ l, respectively¹⁷. The median CD34-positive cell counts on days 4 and 5 in our patients were higher than those reported in this comparative cohort, especially on day 4. The higher values on day 4 might be because we added bortezomib on day 1. We indicate that the addition of bortezomib on both, day 1 and 4, induces PBSC mobilization. We suspended the administration of plerixafor in two patients on the morning of day 4 because we expected to collect enough CD34-positive cells without the treatment. Indeed, we collected adequate cells from both younger age and the better mobilization with bortezomib, administered on day 1, may explain adequate cell collection without plerixafor use.

In conclusion, we found that the combination of bortezomib, G-CSF, and preemptive plerixafor enhanced PBSC mobilization, minimized myeloma cells in the apheresis product, and presented a better objective response than G-CSF and preemptive plerixafor without bortezomib. Moreover, our results indicated that this regimen is reliable and tolerable, as patients experienced only mild toxicities. Thus, we propose a novel autologous PBSC collection regimen comprising bortezomib, G-CSF, and preemptive plerixafor for patients with MM. This method may be preferable to standard protocols, especially for patients who are expected to have a low apheresis yield. Given that we analyzed the results of a small number of patients, further investigation in a larger group of patients is required. Therefore, we are initiating a prospective phase II clinical trial with this mobilization regimen: FBMTG EMM17 (UMIN000028421). We will recruit approximately 50 patients and we expect more reliable results in larger group of patients.

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Author Contributions

K. T. wrote the paper. K. S., I. K., I. Y., and H. H. collected and managed the clinical data.

Conflicts of Interest

The authors declare no conflict of interest. Disclosure forms provided by the authors are available here.

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